

Regulation of genomic DNA methylation by *VeZF1*

Review of: Gowher H, Stuhlmann H, Felsenfeld G. *VeZF1* regulates genomic DNA methylation through its effects on expression of DNA methyltransferase *Dnmt3b*. *Genes Dev* 2008, 22:2075-2084

DNA methylation is one form of epigenetic modification that is responsible for the regulation of gene expression. DNA methylation is maintained throughout a cell's lifespan by de novo DNA methyltransferases *Dnmt3a* and *Dnmt3b* and the maintenance methyltransferase *Dnmt1* (1). Vascular endothelial zinc finger 1 (*VeZF1*) is a recently identified zinc finger transcription factor that is expressed in endothelial cells during vascular development in the mouse embryo (2). *VeZF1*, and its human homologue, DB1, bind in a sequence-specific manner to GC-rich sequences of several genes including metallothionein 1 (2), interleukin-3 (3), endothelin-1 (4), and stathmin/oncoprotein18 (5). Although the mechanism by which *VeZF1* regulates the expression of the DNA methyltransferase *Dnmt3b* is not completely understood, this study shows that the absence of *VeZF1* results in a depletion of *Dnmt3b*, which in turn affects global genomic methylation and the epigenetic regulation of gene expression (6).

Using a mouse embryonic stem cell line in which both copies of *VeZF1* have been deleted (7), Gowher *et al.* present evidence that the absence of *VeZF1* causes major loss of genomic methylation at specific sites, including certain repeat elements, some imprinted loci, and many CpG islands. Using methylated DNA immunoprecipitation (MeDIP), the genome-wide methylation patterns of *VeZF1*^{-/-} embryonic stem cells and wild-type (WT) cells were compared. Following the hybridisation of immunoprecipitated product to UHNMAC Mouse 4.6k CGI arrays, the genome-wide loss of methylation at more than 1300 CpG islands was revealed for *VeZF1*^{-/-} cells. Among these, 76 CGIs were located within 3 Kb upstream of promoters and 71 CGIs were within 3Kb downstream of genes. Many of these CGIs were associated with testis-specific, neuronal-specific, and homeobox genes and genes involved in

tumourigenesis. For 14 of these genes, the expression levels between WT and *VeZF1*^{-/-} were validated using real time-PCR.

Although this study found no significant difference in the overall DNA methylation activity of *VeZF1*^{-/-} and WT cells, differences in the levels of different methylating enzymes were revealed. The expression levels of three known active DNA methyltransferases were compared in *VeZF1*^{-/-} and WT cells. The levels of *Dnmt3a* and *Dnmt1* transcripts were relatively similar, however, *Dnmt3b* expression was considerably reduced in *VeZF1*^{-/-} cells compared with WT. To complicate matters, *Dnmt3b* transcripts are known to undergo alternative splicing and more than eight splice variants have been identified (8). *Dnmt3b1* (full length) and *Dnmt3b6* variants are the major variants of *Dnmt3b* found in normal embryonic stem cells. The mechanism by which *VeZF1* regulates the expression of *Dnmt3b* has not yet been determined, however three binding motifs for *VeZF1* in the introns or 3' UTR of the *Dnmt3b* gene have been identified (6).

This study shows that the partial loss of *Dnmt3b* expression in *VeZF1*^{-/-} embryonic stem cells causes methylation defects similar to those found in ICF (immunodeficiency, centromere instability, facial anomalies) syndrome patients (9,10). ICF syndrome is a genetic disease directly related to a genomic methylation defect that mainly affects classical satellites 2 and 3, both components of constitutive heterochromatin (10). This study shows that *VeZF1*^{-/-} cells display widespread loss of DNA methylation and that this loss correlates with an approximately fourfold reduced expression of the de novo DNA methyltransferase *Dnmt3b*, compared with WT embryonic stem cells (6).

References

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